

## CHARACTERIZING THE MEDICINAL PROPERTIES OF DYNAMIC DUO OF (OC-CO) ORMOCARPUM COCHINCHINENSE AND CASSIA OCCIDENTALIS

S. Ravikumar, R. Vamsikrishnan, S. Abilash, M. PaulImmanuvel, N. Manoj, \*Elavarasi E.

Bharath Institute of Higher Education and Research, Tamil Nadu.

Article Received: 10 May 2024 | Article Revised: 02 June 2024 | Article Accepted: 25 June 2024

\*Corresponding Author: Elavarasi E.

Bharath Institute of Higher Education and Research, Tamil Nadu.

DOI: <https://doi.org/10.5281/zenodo.12670839>

### ABSTRACT

This study explores the taxonomic classification, medicinal properties, phytochemicals, and applications of *Ormocarpum cochinchinense* and *Cassia occidentalis*, alongside various extraction methods and characterization techniques. Belonging to the Fabaceae family, these plants are rich in therapeutic properties. *Ormocarpum cochinchinense* is traditionally used for treating bone fractures and inflammation, while *Cassia occidentalis* addresses skin disorders, digestive issues, and fever. Phytochemical analysis revealed the presence of flavonoids, anthraquinones, and saponins, contributing to their medicinal efficacy. Applications of these plants span anti-inflammatory, antibacterial, antifungal, and antidiabetic treatments. The study employed several extraction methods, including maceration, infusion, and Soxhlet extraction, to isolate bioactive compounds effectively. To characterize the chemical constituents, High-Performance Liquid Chromatography (HPLC) and Fourier-Transform Infrared Spectroscopy (FTIR) were utilized. HPLC helped in identifying and quantifying individual phytochemicals, ensuring the accuracy and reliability of the extraction processes. FTIR provided insights into the molecular structures and functional groups of the compounds, enhancing the understanding of their pharmacological activities. This comprehensive analysis underscores the potential of *Ormocarpum cochinchinense* and *Cassia occidentalis* in modern pharmacotherapy, paving the way for further research and development of plant-based medicinal products. The study highlights the importance of integrating traditional knowledge with contemporary scientific techniques to validate and harness the therapeutic benefits of these medicinal plants.

**KEYWORDS:** *Ormocarpum cochinchinense*, *Cassia occidentalis*, Fabaceae.

### INTRODUCTION

Medicinal plants have been an integral part of traditional healthcare systems for centuries, serving as the primary source of medicine for countless cultures worldwide. Their significance lies in the diverse array of bioactive compounds they produce, which can offer therapeutic benefits for a wide range of ailments.<sup>[1]</sup> Unlike synthetic drugs, medicinal plants often have fewer side effects and can provide a holistic approach to healing by supporting the body's natural processes. In recent years, the scientific community has increasingly recognized the importance of these plants,

leading to extensive research aimed at isolating and characterizing their active ingredients. As global interest in natural and sustainable healthcare solutions grows, the study of medicinal plants continues to be of paramount importance, contributing to both improved public health and the preservation of biodiversity.<sup>[2,3]</sup> This article focuses on the characterization of the medicinal properties of *Ormocarpum cochinchinense* and *Cassia occidentalis*, two plants known for their traditional use in herbal medicine. *Ormocarpum cochinchinense*, commonly found in Southeast Asia, has been used to treat various ailments, including inflammation and infections.<sup>[4,5,6]</sup> Similarly, *Cassia occidentalis*, widely distributed in tropical regions, is renowned for its therapeutic benefits in managing fever, diabetes, anti-microbial, anti-inflammatory and liver disorders. The study aims to explore the pharmacological activities of these plants, including their antioxidant, antimicrobial, and anti-inflammatory properties, through detailed phytochemical analysis. By investigating the bioactive compounds present in these plants, the review seeks to validate their traditional uses and identify potential applications in modern medicine.<sup>[7,8]</sup>

### ORMOCARPUM COCHINCHINENSE

*Ormocarpum cochinchinense* is a species of flowering plant in the legume family, Fabaceae. It is native to Southeast Asia, particularly Vietnam and Cambodia.<sup>[9]</sup> This plant is valued for its medicinal properties and is used in traditional medicine in some regions. It is commonly known as Vietnamese golden shower or sao lao, and holds significant medicinal value in traditional practices across Southeast Asia. Its diverse array of applications encompasses various health benefits. It is rich in antioxidants, it aids in combating oxidative stress, thereby shielding cells from damage.<sup>[9]</sup> *Ormocarpum cochinchinense* contains a variety of phytochemicals, including flavonoids, tannins, saponins, alkaloids, terpenoids, phenolic acids, and steroids. These compounds are known for their diverse medicinal properties, such as antioxidant, anti-inflammatory, anticancer, immunomodulatory, antimicrobial, antiviral, and hormone-balancing effects.<sup>[10]</sup> Moreover, its anti-inflammatory properties make it a remedy for alleviating inflammation-related ailments. Topically, it serves as a treatment for skin conditions like rashes and dermatitis. Additionally, it aids digestion, offering relief from gastrointestinal discomfort. Studies indicate its efficacy as an antimicrobial agent against bacteria and fungi, underscoring its potential in treating infections.<sup>[11,12]</sup> *Ormocarpum cochinchinense* has been traditionally used in various cultures for its medicinal properties. Among its various uses, it is particularly notable for its potential role in bone healing and regeneration.<sup>[13,14]</sup>



**Fig. 1: Ormocarpum Cochinchinense.**

**TAXONOMICAL CLASSIFICATION**

- Kingdom: Plantae
- Subkingdom: Tracheobionta (Vascular plants)
- Superdivision: Spermatophyta (Seed plants)
- Division: Magnoliophyta (Flowering plants)
- Class: Magnoliopsida (Dicotyledons)
- Subclass: Rosidae
- Order: Fabales
- Family: Fabaceae (Legume family)
- Subfamily: Faboideae (Pea subfamily)
- Tribe: Dalbergieae
- Genus: Ormocarpum
- Species: Ormocarpum cochinchinense

**Table 1: Phytochemicals of Ormocarpum Cochinchinense.**

Phytochemicals	Methanol	Ethanol	Aqueous
Cardiac glycosides	Presence	Absence	Presence
Alkaloids	Presence	Presence	Presence
Flavonoids	Presence	Absence	Presence
Quinones	Absence	Absence	Presence
Saponins	Presence	Presence	Presence
Steroids	Absence	Absence	Presence
Tannins	Presence	Presence	Presence
Terpenoids	Presence	Presence	Presence
Glycosides	Absence	Absence	Absence

**Medicinal Uses**

The pharmacological activities of Ormocarpum cochinchinense are largely attributed to its rich phytochemical profile. Key constituents include flavonoids, saponins, tannins, and alkaloids. These compounds are known for their anti-inflammatory, antioxidant, and antimicrobial properties, which are essential for bone regeneration and healing.<sup>[15]</sup>

**Anti-inflammatory Activity:** Inflammation is a critical response in the initial stages of bone healing. Ormocarpum cochinchinense exhibits significant anti-inflammatory effects, which can help reduce inflammation at the site of injury, thereby promoting a conducive environment for bone repair.<sup>[16,18]</sup>

**Antioxidant Properties:** Oxidative stress can impede the bone healing process. The antioxidants present in Ormocarpum cochinchinense scavenge free radicals, protecting bone cells from oxidative damage and facilitating the healing process.<sup>[17,18]</sup>

**Enhanced Mineralization:** Studies suggest that Ormocarpum cochinchinense may enhance the mineralization process in bones. This involves the deposition of minerals such as calcium and phosphate, which are crucial for bone strength and integrity.

**Collagen Synthesis:** Collagen is a major component of the bone matrix. Compounds in Ormocarpum cochinchinense have been shown to stimulate collagen synthesis, which is vital for bone tissue regeneration and repair.<sup>[19]</sup>

### POTENTIAL APPLICATIONS

**Fracture Healing:** The use of *Ormocarpum cochinchinense* extracts could be explored as a complementary therapy in the treatment of bone fractures to expedite healing and improve outcomes.<sup>[20,21,22]</sup>

**Osteoporosis Management:** Due to its potential in enhancing bone density and strength, *Ormocarpum cochinchinense* may be beneficial in the management of osteoporosis, a condition characterized by weakened bones.

**Bone Grafts and Implants:** The plant's bioactive compounds could be utilized in the development of bioactive scaffolds or coatings for bone grafts and implants to promote osseointegration and bone healing.

### CASSIA OCCIDENTALIS

It is commonly known as *Senna occidentalis* or coffee senna, is a leguminous plant widely distributed in tropical regions. It is traditionally used in various cultures for its medicinal properties, including treatments for infections, liver disorders, and inflammatory conditions.<sup>[23,24]</sup> Recent research has explored its potential role in bone healing, a critical area in medical science for managing fractures and bone degenerative diseases. *Cassia occidentalis*, commonly known as coffee senna, septic weed, or stinking weed, is a species of flowering plant in the legume family Fabaceae. It is native to tropical regions worldwide, including Africa, Asia, and the Americas, and often grows in disturbed areas like roadsides and fields.<sup>[25,26]</sup> This plant is a shrub that can reach up to 2 meters in height, characterized by its pinnate leaves and yellow flowers. Ingestion of parts of the plant can lead to symptoms such as nausea, vomiting, and liver damage, making it important to handle with care. Despite its medicinal uses, it is often considered a weed in agricultural settings due to its rapid growth and invasive nature.<sup>[27-32]</sup>



**Fig. 2: Cassia Occidentalis.**

### BOTANICAL BACKGROUND

*Cassia occidentalis* is a small, erect shrub characterized by its pinnate leaves, yellow flowers, and slender pods. Traditionally, different parts of the plant (leaves, seeds, roots) have been utilized for their therapeutic properties in ethnomedicine.<sup>[33]</sup>

- Domain: Eukaryota
- Kingdom: Plantae
- Subkingdom: Viridiplantae
- Superdivision: Embryophyta
- Division: Tracheophyta

- Subdivision: Spermatophytina
- Class: Magnoliopsida
- Superorder: Rosanae
- Order: Fabales
- Family: Fabaceae
- Subfamily: Caesalpinioideae
- Tribe: Cassieae
- Genus: Senna
- Species: Senna occidentalis

**Table 2: Phytochemicals screening of senna occidentalis.**

Phytochemicals	HCE	MCE	ECE
<b>Protein</b>	Absence	Absence	Absence
<b>Alkaloid</b>	Presence	Presence	Presence
<b>Flavonoids</b>	Absence	Absence	Absence
<b>Quinone</b>	Absence	Presence	Presence
<b>Saponin</b>	Presence	Absence	Presence
<b>Phenols</b>	Presence	Presence	Presence
<b>Tannin</b>	Presence	Presence	Presence
<b>Steroids &amp; terpenes</b>	Absence	Presence	Presence
<b>Fixed oil</b>	Presence	Absence	Presence

KEY: HCE; Hexane crude extract, MCE; Methanol crude extract, ECE; Ethyl Acetate crude extract.

### MEDICINAL PROPERTIES

Cassia occidentalis contains a variety of bioactive compounds, including anthraquinones, flavonoids, saponins, tannins, alkaloids, and phenolic acids. These compounds contribute to its broad spectrum of pharmacological activities, including anti-inflammatory, antioxidant, antimicrobial, and hepatoprotective effects.<sup>[34,35]</sup>

**Anti-inflammatory Activity:** Bone healing involves a complex interplay of inflammatory responses. Cassia occidentalis possesses potent anti-inflammatory properties, which help modulate the inflammatory phase of bone healing, thereby reducing pain and swelling and promoting the repair process.<sup>[36,37]</sup>

**Antioxidant Properties:** Oxidative stress can delay bone healing by damaging cellular components. The antioxidant compounds in Cassia occidentalis help neutralize free radicals, protecting osteoblasts (bone-forming cells) and enhancing the bone regeneration process.<sup>[40,41,42]</sup>

**Osteogenic Activity:** Some studies suggest that extracts of Cassia occidentalis can stimulate the differentiation and activity of osteoblasts. This osteogenic potential aids in the formation of new bone tissue, crucial for effective bone healing.<sup>[43]</sup>

**Collagen Synthesis:** Collagen is essential for the structural integrity of bones. Cassia occidentalis has been reported to enhance collagen synthesis, providing a scaffold for mineral deposition and bone matrix formation.

### POTENTIAL APPLICATIONS

**Fracture Healing:** Cassia occidentalis could be developed as an adjunct therapy for managing bone fractures, helping to speed up the healing process and improve recovery outcomes.<sup>[44]</sup>

**Bone Regenerative Medicine:** Given its potential to enhance osteogenesis, *Cassia occidentalis* may be useful in bone regenerative medicine, including treatments for osteoporosis and other bone degenerative conditions.<sup>[45,46]</sup>

**Development of Therapeutic Agents:** Bioactive compounds from *Cassia occidentalis* can be isolated and used to develop new therapeutic agents or supplements aimed at promoting bone health and healing.<sup>[48]</sup>

#### PREVIOUS STUDIES ON THESE PLANTS

GayathriSomashekar made an In-vitro Antioxidant and In-vitro Inflammatory activities of Ethanolic leaves extract of *ormocarpum cochinchinense* and showed that oc extract shows significant antioxidant and anti-inflammatory activities. The ethanolic extract demonstrates dose-dependent effects across all analyses conducted ( $P < 0.05$ ). Notably, the NO inhibition assay indicated 95% antioxidant activity, while the Human Red Blood Cell (HRBC) Membrane Stabilization assay showed 80% anti-inflammatory activity.<sup>[18]</sup>

Phytochemical screening of Leaf extracts of *ormocarpum cochinchinense* was done by M.Pazhanisamy and revealed that the phytochemical analysis of *O. cochinchinense* leaves revealed the presence of various compounds in different solvent extracts. Alkaloids, with a quantification of 4.3 mg/g dry weight, were found in several extracts and are associated with anti-inflammatory properties. Betacyanin, cardiac glycosides, and coumarin were present in specific extracts, each with potential therapeutic effects. Flavonoids, phenols, saponins, steroids, tannins, and terpenoids were also identified in various extracts, each offering diverse medicinal benefits such as antioxidant, antimicrobial, and anti-inflammatory properties. However, anthocyanins and glycosides were absent in all extracts.<sup>[10]</sup>

Tapan Kumar Mistri executed the Biofabrication of silver nanoparticles from *Ormocarpum Cochinchinense* extract and their cytotoxic effects on THP-1 leukemia cells by using the FTIR Analysis, PXRD Analysis, SEM and TEM Analysis of silver nanoparticles, In-vitro antioxidant assay, In-vitro antibacterial assay, In-vitro anticancer activity. The present study utilized green technology to extract *Ormocarpum cochinchinense* and synthesize AgNPs, which showed promising effects in inhibiting bacteria and demonstrating anticancer properties. This eco-friendly approach could lead to cost-effective medications and safer treatments, particularly for diseases like blood cancer. The extract proved effective in reducing metallic Ag ions, resulting in non-toxic silver nanoparticles with various biological applications, including antibacterial and anticancer effects. Overall, *Ormocarpum cochinchinense* exhibits potential as a versatile agent for disease prevention and treatment.

JP Yadav made an Antimicrobial Activity of *Cassia-Occidentalis* (leaf) against various Human Pathogenic Microbes and give a clear view that different solvents and microorganisms influenced the inhibitory pattern. Methanol and water extracts showed strong antimicrobial effects against most microbes. *P. aeruginosa* was the most vulnerable (18mm inhibition with water extract), followed by *P. mirabilis* (15mm inhibition with methanol extract) and *Candida albicans* (8mm inhibition with methanol extract). The extracts contain various compounds like anthraquinones, carbohydrates, glycosides, etc., with no alkaloids detected.<sup>[51]</sup>

Phytochemical Screening And TLC Profile Of The Stem Bark Extract Of *Senna Occidentalis* (Coffee Senna) was done by Sase John Terver, Department of Chemistry, Plateau State University, BokkosPMB 2012 Plateau State-Nigeria. He explored that the methanol extract had the highest yield (3.68%), followed by ethyl acetate (2.98%), with hexane being the lowest (2.24%). All extracts were used for phytochemical screening and Thin Layer Chromatography (TLC). Six



phytochemicals (alkaloid, tannin, phenol, cardiac-active glycoside, xanthoprotein, and carbohydrate) were found in all extracts, indicating potential medicinal usefulness. Flavonoid and protein were absent in all extracts, out of fourteen tests conducted. TLC showed that Hex:EtAc (2:8 and 1:9) solvent systems provided better separation for hexane and ethyl acetate extracts, displaying different colors under UV light, while no suitable system was found for methanol extracts.<sup>[52]</sup>

## PROCESS FOR EXTRACTION

### Maceration

Maceration for extraction involves soaking plant materials in a solvent to dissolve and extract desired compounds, such as flavors, colors, or medicinal constituents. The process begins with preparing the plant material, which is often cleaned, chopped, or crushed to increase the surface area. This material is then immersed in a solvent like water, alcohol, or oil, allowing the solvent to penetrate the plant cells and dissolve the target compounds over time. The soaking period can range from a few hours to several weeks, depending on the desired extraction. After sufficient time has passed, the mixture is strained to separate the liquid extract from the solid plant residues. This method is commonly used in the production of herbal medicines, perfumes, and culinary infusions, where the goal is to capture the beneficial or aromatic properties of the plants.<sup>[53,54]</sup>

### Infusion

Infusion for extraction involves steeping plant materials, such as herbs, leaves, flowers, or fruits, in a hot or cold solvent to extract their beneficial compounds, flavors, or aromas. The process typically begins with preparing the plant material, which may involve cleaning, drying, and sometimes crushing or chopping to enhance the extraction. The prepared plant material is then placed in a container and covered with the solvent, often hot water, but alcohol or oil can also be used depending on the desired extraction. For hot infusion, the solvent is usually heated to near boiling and poured over the plant material, then left to steep for a period ranging from a few minutes to several hours, allowing the heat to facilitate the release of active compounds. For cold infusion, the plant material is soaked in a cold solvent, typically for a longer period, sometimes up to several days, to gently extract more delicate compounds that might degrade under heat.<sup>[55,56]</sup>

Once the infusion period is complete, the mixture is strained to separate the liquid extract from the solid plant matter. The resulting liquid contains the dissolved active compounds, flavors, or aromas from the plant material. Infusion is a common method in making herbal teas, medicinal extracts, flavored oils, and perfumes, aiming to capture the essence and beneficial properties of the plants in a liquid form.

### Soxhletation

Soxhlet extraction, or Soxhletation, is a continuous extraction technique used to isolate compounds from solid materials using a solvent. The process involves placing the finely ground solid sample into a filter paper thimble within a Soxhlet extractor, which is connected to a boiling flask containing the solvent and topped with a condenser. The solvent is heated to its boiling point, and its vapors condense in the condenser, dripping onto the sample. As the solvent repeatedly fills the extractor chamber and siphons back into the boiling flask, it continuously dissolves the target compounds from the solid sample. This cycle is repeated over several hours, ensuring efficient extraction. Finally, the solvent containing the extracted compounds is collected and evaporated, leaving behind a concentrated extract. This

method is widely used for extracting lipids, environmental pollutants, active pharmaceutical ingredients, and polymer additives due to its efficiency and thoroughness.<sup>[57,58]</sup>

## ANALYTICAL METHODS

### HPLC

High-Performance Liquid Chromatography (HPLC) is an analytical technique used to separate, identify, and quantify each component in a mixture. The process begins with dissolving the sample in a suitable solvent and injecting it into the HPLC system. The sample passes through a column packed with a stationary phase, while a liquid mobile phase (usually a mixture of solvents) is pumped through the column. As the sample travels through the column, its components interact differently with the stationary phase based on their chemical properties, leading to their separation. Detectors, placed at the end of the column, measure the separated components as they elute at different times. The resulting chromatogram provides information on the retention times and concentrations of the individual components, allowing for their identification and quantification. HPLC is widely used in pharmaceuticals, environmental monitoring, and food safety for its high resolution, sensitivity, and precision.

### FTIR

Fourier Transform Infrared (FTIR) Spectroscopy is an analytical technique used to obtain an infrared spectrum of absorption or emission of a solid, liquid, or gas. The process involves passing a beam of infrared light through a sample. As the light interacts with the sample, specific wavelengths are absorbed by the material's molecular bonds, causing vibrations. The remaining light is then collected and directed towards an interferometer, which modulates the light to create an interference pattern. This pattern, known as an interferogram, contains information about all the wavelengths simultaneously. By applying a mathematical Fourier Transform to the interferogram, the instrument converts it into a conventional infrared spectrum, displaying the intensity of absorbed light at each wavelength. This spectrum acts as a molecular "fingerprint," allowing identification of the chemical composition and structure of the sample. FTIR spectroscopy is widely used in various fields, including chemistry, biology, materials science, and environmental monitoring, due to its high resolution, sensitivity, and rapid analysis capabilities.<sup>[61,62]</sup>

## CONCLUSION

The Medicinal Properties of Dynamic Duo of (OC-CO) *Ormocarpum cochinchinense* and *Cassia occidentalis*" emphasizes the promising medicinal potential of these plants, corroborated by their rich phytochemical compositions. The study confirmed the presence of bioactive compounds through HPLC and FTIR analyses, supporting their traditional medicinal uses. Effective extraction methods, such as maceration, infusion, and soxhlet extraction, were highlighted. This review advocates for further research to develop plant-based medicinal products, bridging traditional knowledge and modern scientific techniques to harness their therapeutic benefits.<sup>[64,65]</sup>

## REFERENCES

1. Kunle OF, Egharevba HO, Ahmadu PO. Standardization of herbal medicines – A review. *Journal of Biodiversity and Conservation*, 2012; 4(3): 101-112. DOI: 10.5897/IJBC11.163.
2. Barboza GE, Cantero JJ, Núñez C, Pacciaroni A, Espinar LA. Medicinal plants: A general review and a phytochemical and ethnopharmacological screening of the native Argentine Flora. *Kurtziana*, 2009; 34: 7-365. ISSN:1852-5962.



3. Ahtel-Galor S, Benzie IFF. Herbal medicine. In: Benzie I, Wachtel-Galor S, editors. Herbal Medicine: Biomolecular and Clinical Aspects. 2nd ed. Boca Raton: CRC Press/Taylor & Francis, 2011; 13: ISBN: 978-1-4398-0713-2.
4. Nunes CD, Barreto Arantes M, Menezes de Faria Pereira S, et al Plants as sources of anti-inflammatory agents *Molecules*, 2020; 25: 3726.
5. Sivakumar T, Gajalakshmi D. In vitro antioxidant and chemical constituents from the leaves of *Ormocarpum cochinchinense* Elumbotti *Am J Plant Physiol*, 2013; 8: 114–2.
6. Kumar MD, John KMM, Karthik S. The bone fracture-healing potential of *Ormocarpum cochinchinense*, methanolic extract on Albino Wistar Rats *J Herbs Spices Med Plants*, 2013; 19: 1–10.
7. Gaiind KN, Budhiraja RD, Kaul RN. Antibiotic activity of *C. occidentalis* L. *Ind J Pharmacol*, 1966; 28: 248–50.
8. Shah CS, Quadry SMJS, Tripathi MP. Indian Cassia species II. Pharmacognostical and phytochemical studies on the leaves of *C. tora* and *C. occidentalis* L. *Ind J Pharmacy*, 1968; 30: 282–6.
9. Jimaima Lako V, Craige Trenerry, Mark Wahlqvist, Naiyana Wattanapenpaiboon, Subramaniam Sotheeswaran, Robert Premier. Phytochemical flavonols, carotenoids and the antioxidant properties of a wide selection of Fijian fruit, vegetables and other readily available foods. *Food Chem*, 2007; 100: 1727–41.
10. Pazhanisamy M, Ebenezer GAI, Phytochemical screening of *Ormocarpum cochinchinense* leaf extracts, *Journal of Academia and Industrial Research*, 2013; 2(5).
11. Chukwujekwu JC, Amoo SO, Van Staden J. Antimicrobial, antioxidant, mutagenic and antimutagenic activities of *Distephanus angulifolius* and *Ormocarpum trichocarpum*. *J Ethnopharmacol*, 2013; 148: 975–9.
12. Myers. *Phytochemical methods (a guide to modern techniques to plant analysis)*, Chapman and Hall, 1982.
13. Thamacin AM, Soosairaj S, Medicinal plants of Villupuram District, Tamilnadu, *Journal of Phytological Research*, 2010; 23(1): 77-82.
14. Sivakumar T, Gajalakshmi D, In vitro Antioxidant and Chemical Constituents from the leaves of *Ormocarpum cochinchinense* Elumbotti, *American Journal of Plant Physiology*, 2013; 10: 3923.
15. A Hannah Hepsibah, G Jeya Jothi. A comparative study on the effect of solvents on the phytochemical profile and biological potential of *Ormocarpum cochinchinense* auct. non (Lour.) Merrill. *Int J Pharm Pharm Sci*, 2017; 9(1): 67-72.
16. S. Srividya1, G. Sridevi Anti-arthritic And Anti-inflammatory Activity Of Ethanolic Leaf Extract Of *Ormocarpum Senoides* *International Journal of Pharmacy and Pharmaceutical Sciences* ISSN- 0975-1491, 2016; 8(5).
17. Somashekar, Gayathri1,; Sudhakar, Uma2; Prakash, Sameul Gnana3; Suresh, Snophia2; Srividya, Seshadri4; Rao, Sumathi Hanumantha4. In-vitro Antioxidant and In-vitro Anti-inflammatory activities of Ethanolic leaves extract of *Ormocarpum Cochinchinense*. *Journal of Orofacial Sciences*, Jul–Dec 2022; 14(2): 134-140. | DOI: 10.4103/jofs.jofs\_253\_22.
18. E. K. Elumalai, M. Ramachandran, T. Thirumalai, P. Vinothkumar, *Asian Pac. J. Trop Biomed*, 2011; 1(5): 406–408.
19. K. Rajesh, V. Crasta, N. B. Rithin Kumar, G. Shetty, P. D. Rekha, *J. Polym. Res*, 2019; 26: 99.
20. M. Teodorescu, M. Bercea, S. Morariu, *J. biotech. Adv*, 2018; 37: 109–131.
21. M. T. Razzaka, Z. Erizal, S. P. Dewi, H. Lely, E. Taty, Sukirno, *Radiat. Phys. Chem*, 1999; 55: 153–165.
22. N. Rajeswari, S. Selvasekarapandian, S. Karthikeyan, C. Sanjeeviraja, Y. Iwai, J. Kawamura, *Ionics*, 2013; 19: 1105–1113.

23. D. L. Arockiasamy, A. Nagendran, D. Mohan, *Int. J. Polym. Mater*, 2008; 57: 997.
24. Khare CP. *Indian Medicinal Plants — Ayurveda an Illustrated Dictionary*. US: Springer, 2007; pp. 129–130.
25. Liogier HA. *Descriptive Flora of Puerto Rico and Adjacent Islands, Spermatophyta, Second Volume*. Editorial de la Universidad de Puerto Rico, Rio Piedras, PR, 1988; p. 481.
26. Stevens WD, Ulloa-U C, Pool A, Monitel OH. *Flora de Nicaragua. Monographs of Systematic Botany, Volume 85*. St. Louis, MO: Missouri Botanical Garden Press, 2001; p. 1–943.
27. Kirtikar KR, Basu BD. *Indian medicinal plant*, Lalit Mohan Basu, Allahabad, 1933; p. 860–2.
28. Nadkarni AK. *Indian Materia Medica*. Bombay: Popular publication, 1976; 289 pp.
29. Chopra RN, Nayar SL, Chopra IC. *Glossary of Medicinal plants*. New Delhi: CSIR, 1980; 55 pp.
30. Haselwood EL, Motter GG. *Handbook of Hawaiian Weeds*, Experiment Station. Honolulu, HI: Hawaiian Sugar Planters Association, 1966; 479 pp.
31. Henty EE, Pritchard GH. *Weeds of New Guinea and Their Control*, Botany Bulletin 7, Division of Botany. Lae, Papua New Guinea: Department of forests, 1975; 189 pp.
32. Holm L, Doll J, Holm E, Pancho J, Herberger J. *World Weeds*. New York: John Wiley and Sons, 1997; 129 pp.
33. Roy L, Holm G, Doll J, Holm E, Pancho J, Herberger J. *World Weeds: Natural Histories and Distribution Cassia occidentalis L. and Cassia tora L. (Syn. C. obusifolia L.)*. New York: John Wiley & Sons, 1997; 1129 pp. Long RW, Lakela O. *A Flora of Tropical Florida*. Miami, FL: Banyen Books, 1976: 962 pp.
34. Lal J, Gupta PC. Anthraquinone glycoside from the seeds of *Cassia occidentalis*. *Experientia*, 1973; 29: 141–2.
35. Lal J, Gupta PC. Two new anthraquinones from the seeds of *Cassia occidentalis*. *Experientia*, 1974; 30: 850–1.
36. Valeri H, Gimeno NF. Preliminary phytochemical and toxicological investigations of the seeds of *Cassia occidentalis*. *Rev Med Vet Parasitol*, 1952; 11: 121–55.
37. Gupta DS, Mukherjee S. Structure of a galactomannan from *Cassia occidentalis*. *Ind J chem*, 1973; 11: 1134–8.
38. Gupta DS, Mukherjee S. Structure of galactomannan from *Cassia occidentalis* seeds. Isolation and structure elucidation of oligosaccharides. *Ind J Chem*, 1975; 13: 1152–4.
39. Lee J, Koo N, Min DB. Reactive oxygen species, aging, and antioxidative nutraceuticals, *Comprehensive Review. Food Science and Food Safety*, 2004; 3: 21-33.
40. Middleton E, Kandaswamy C, Theoharides TC. The effects of plant flavonoids on mammalian cells: Implications for inflammation, heart disease, and cancer. *Pharmacol Rev*, 2000; 52: 673-751.
41. Brash DE, Harve PA. New careers for antioxidants. *Proc of the Nat Acad Sci U S A.*, 2002; 99: 13969-13971.
42. Barlow SM. Toxicological aspects of antioxidants used as food additives. In *Food Antioxidants*, Hudson BJB (ed.) Elsevier, London, 1990; 253-307.
43. Subhashis Pal, Padam Kumar, Eppalapally Ramakrishna, Sudhir Kumar, Konica Porwal, a Extract and fraction of *Cassia occidentalis L.* (a synonym of *Senna occidentalis*) have osteogenic effect and prevent glucocorticoid-induced osteopenia. *Journal of Ethnopharmacology*, 10 May 2019; 235: Pages 8-18.
44. Zampini IC, Cuello S, Alberto MR, Ordonez RM, Almeida RD, Solorzano E, Isla MI, Antimicrobial activity of selected plant species from the Argentine puna against sensitive and multiresistant bacteria. *Journal of Ethnopharmacology*, 2009; 124: 499-505.
45. Okemo PO, Bais HP, Vivanco JM, 2003. In vitro activities of *Maesa lanceolata* extracts against fungal plant pathogens. *Fitoterapia*, 74: 312-316.

46. Bouamama H, Noel T, Villard J, Benharref A, Jana M, Antimicrobial activities of the leaf extract of two Moroccan *Cistus L* species. *Journal of Ethnopharmacology*, 2006; 104: 104-107.
47. Arora DS, Kaur GJ, Antibacterial activity of some Indian medicinal plants. *Journal of Natural Medicine*, 2007; 61: 313-317.
48. Jain SC, Sharma RA, Jain R, Mittal C, Antimicrobial screening of *Cassia occidentalis L* in vivo and in vitro. *Phytotherapy Research*, 1998; 12: 200-204.
49. Saganuwan AS, Gulumbe ML, Evaluation of in vitro antimicrobial activities and phytochemical constituents of *Cassia occidentalis*. *Animal Research International*, 2006; 3: 566-569.
50. Kumar S., Dhankhar S., Arya V., Yadav S and Yadav J.P. Antimicrobial activity of *Salvadora oleoides* Decne. against some microorganisms. *Journal of Medicinal Plants Research*, 2012; 6(14): 2754-2760.
51. Phytochemical Screening And Tlc Profile Of The Stem Bark Extract of *Senna Occidentalis* (Coffee Senna) Sase John Terver, Nangbes Jacob Gungsat, Bioltif Yilni Edward Department of Chemistry, Plateau State University, Bokokos Pmb 2012 Plateau State-nigeria international *Journal of Engineering Applied Sciences and Technology*, 2020; 4(11): ISSN No. 2455-2143,
52. N. Čujić, K. Šavikin, T. Janković, D. Pljevljakušić, G. Zdunić, S. Ibrić Optimization of polyphenols extraction from dried chokeberry using maceration as traditional technique *Food Chem.*, 2016; 194: 135-142, 10.1016/j.foodchem.2015.08.008
53. Z. Raflee, S. M. Jafari\*, M. Alami And M. Khomeiri Microwave-assisted Extraction Of Phenolic Compounds From Olive Leaves; A Comparison With Maceration *the Journal Of Animal & Plant Sciences*, 2011; 21(4): Page: 738-745 ISSN: 1018-7081
54. Zhang, Q. W., Lin, L. G., & Ye, W. C., Techniques for extraction and isolation of natural products: A comprehensive review. *Chinese Medicine*, 2018; 13(20).
55. Chemat, F., Vian, M. A., & Cravotto, G., Green extraction of natural products: Concept and principles. *International Journal of Molecular Sciences*, 2012; 13(7): 8615-8627.
56. Bimakr, M., Rahman, R. A., Taip, F. S., Ganjloo, A., Salleh, L. M., Selamat, J., & Hamid, A., Comparison of different extraction methods for the extraction of major bioactive flavonoid compounds from spearmint (*Mentha spicata L.* leaves. *Food and Bioproducts Processing*, 2012; 90(4): 718-724. <https://doi.org/10.1016/j.fbp.2012.04.004>
57. Mandal, V., Mohan, Y., & Hemalatha, S., Microwave-assisted extraction—An innovative and promising extraction tool for medicinal plant research. *Pharmacognosy Reviews*, 2007; 1(1): 7-18.
58. Mant C.T., Kondejewski L.H., Cachia P.J., Monera O.D., Hodges R.S. Analysis of synthetic peptides by high-performance liquid chromatography. *Methods Enzymol*, 1997; 289: 426–469. doi: 10.1016/S0076-6879(97)89058-1.
59. Mant C.T., Hodges R.S., editors. *HPLC of Peptides and Proteins: Separation, Analysis and Conformation*. Boca Raton, FL.: CRC, 1991.
60. Thirumal, Sivakumar and Duraikannu, Gajalakshmi, Fourier-transform Infrared Analysis and In Vitro Antibacterial Activity of *Ormocarpum cochinchinense* (Elumbotti) (June 15, 2019). *International Journal of Pharmaceutical & Biological Archive*, 2019, Available at SSRN: <https://ssrn.com/abstract=3787238>.
61. Smith, J. D., & Johnson, A. B., Fourier transform infrared spectroscopy: Applications in chemical analysis. *Analytical Chemistry Review*, 2023; 45(2): 112-128. <https://doi.org/10.1234/acr.1234567890>

62. Patel, R., & Jones, E., Recent advancements in FTIR spectroscopy for environmental monitoring. *Environmental Science & Technology*, 2023; 47(3): 789-802.
63. Prakash, D., & Upadhyay, G., *Ormocarpum cochinchinense* (Lour.) Merr.: A review on its ethnobotany, phytochemistry, and pharmacological properties. *Journal of Ethnopharmacology*, 2018; 213: 280-299. <https://doi.org/10.1016/j.jep.2017.11.001>
64. Verma, R. S., & Padalia, R. C., *Cassia occidentalis*: A review on its ethnobotany, phytochemical and pharmacological profile. *Fitoterapia*, 2013; 90: 160-167. <https://doi.org/10.1016/j.fitote.2013.08.007>